

HERMOSILLOL, AN 8,3'-NEOLIGNAN FROM *KRAMERIA SONORAE**

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Key Word Index—*Krameria sonorae*; Krameriaceae; hermosillo; 8,3'-neolignan.

Abstract—The aerial parts of *Krameria sonorae* afforded seven known neolignans and nor-neolignans, and as a major component, a new 8,3'-neolignan which was named hermosillo.

INTRODUCTION

Krameria species occur in arid areas of tropic and subtropic America [2–4]. Plants of the genus *Krameria* were originally considered to be part of the Leguminosae but later were established as an individual botanical family [5]. In the course of our investigations on the non-polar constituents of *Krameria* species we have studied *K. sonorae* Britt. [3] plants collected in Northern Mexico.

RESULTS AND DISCUSSION

The aerial parts of *K. sonorae* afforded seven known compounds (1–7) [1, 6–9] and, as a major component, the new compound 8.

The ¹H NMR spectrum of 8 contains signals which are attributable to two phenyl systems, one of which is *p*-hydroxy substituted, the other methoxy and propenyl substituted. In addition, it exhibits signals of an ABX₂-system at δ 5.01 (1H, *m*), 5.09 (1H, *m*) and 3.69 [2H, *s* (*br*)]. Irradiation at δ 3.69 transforms both multiplets into doublets ($J = 2.0$ Hz) thus establishing the presence of partial structure a.

Location of the hydroxyl group at ring A is proved by the MS of 8 and its hydrogenation product 9, which both show characteristic fragments at m/z 107 (after H/D-exchange: m/z 107 $\rightarrow$ 108) and, in the case of 9, a base peak at m/z 177 (no shift on H/D-exchange).

Ring B according to the ¹H NMR data is 1,2,5-substituted. The positions of the individual substituents are given by (i) increment calculations using ¹³C NMR data (see Table 1) and (ii) the effect caused by the OCH₃-substituent on the ¹³C-resonance of C-8 in 9. By comparison with the data published for aurein [10, 11] this shift was found to be $\Delta\delta_{C-8} = -7.5$ ppm.

Although neolignans with a 1,2-diphenyl-propane (= lancifolin type) skeleton are found in nature [12, 13], hermosillo (8) is the first compound of the hitherto unknown 1,2-diphenyl-2-propene type.

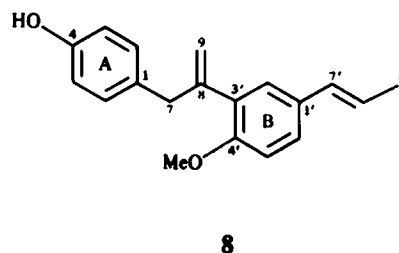
EXPERIMENTAL

Plant material. Plants were collected around Hermosillo, Sonora (Mexico) in November 1985; a voucher specimen is kept at our herbarium (Reg. No. 8032) in Monterrey.

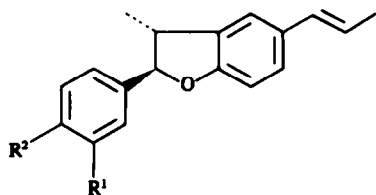
Extraction and chromatography. Aerial parts (370 g) were extracted successively with hexane and MeOH to yield 2.2 g hexane extract and 16.0 g MeOH extract. The hexane extract was repeatedly chromatographed on silica gel (Macherey-Nagel 815338) using: cyclohexane–EtOAc, CHCl₃–MeOH and CH₂Cl₂ (at 7°). This yielded compounds 1–8.

General procedures. TLC: Nano plates Sil-20 UV (Macherey-Nagel), solvent systems: S-1 = cyclohexane–EtOAc (17:1), S-2 = CHCl₃, detection by UV and anisaldehyde reagent [14] followed by heating; IR: CHCl₃; UV: MeOH, unless stated otherwise; ¹H NMR: 90 and 400 MHz, CDCl₃; ¹³C NMR: 22.5 MHz, CDCl₃, int. reference TMS; MS: 70 eV.

Identifications of compounds 1 to 7 were based on comparisons with authentic substances obtained from *K. cystioides* according to [1].

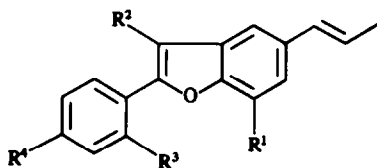


* Part 2 in the series 'Studies on Krameriaceae'. For part 1 see ref. [1].

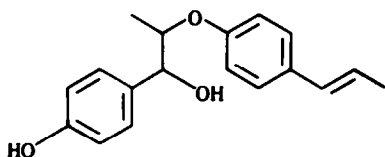


(Absolute configuration)

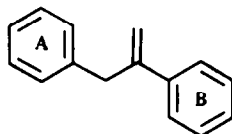
	R ¹	R ²
1	H	OH (= conocarpan)
2	OMe	



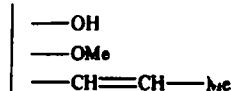
	R ¹	R ²	R ³	R ⁴
3	H	Me	H	OH (= eupomatenoid 6)
4	OMe	Me	H	OH (= eupomatenoid 13)
5	H	H	OH	OMe (= ratanhia-phenol I)
6	H	H	OH	OH



(Relative configuration) 7



8



Hermosillo (8) (4-[2-(2-Methoxy-5-(*E*)-propenyl)-phenyl]allylphenol. Oil (73 mg); TLC (S-1): R_f 0.28, violet with anisaldehyde; IR $\nu_{\max}$ cm^{-1} : 3590 (OH), 2930, 2855, 2840, 1600; UV $\lambda_{\max}$ (log ϵ): 226(4.51), 262(4.49), 270(4.47, sh), 287(4.09, sh), 297(3.94); + NaOH: 220–262 (4.52, plateau), 272(4.47, sh), 295(4.38); ^1H NMR (400 MHz): δ 1.83 (3H, *d* (*br*), J = 5 Hz, CH_3 -9'), 3.69 (2H, *s* (*br*), CH_2 -7), 3.82 (3H, *s*, MeO), 4.70 (1H, *s*, (*br*), OH), 5.01 (1H, *m*, H_A -9), 5.09 (1H, *m*, H_B -9), 6.06 (1H, *dq*, J_1 = 16, J_2 = 5 Hz, H-8'), 6.31 (1H, *d* (*br*), J = 16 Hz, H-7'), 6.71 (2H, AA'BB', J = 8.5 Hz, H-3, H-5), 6.78 (1H, *d*, J = 8.5 Hz, H-5'), 7.03 (2H, AA'BB', J = 8.5 Hz, H-2, H-6), 7.05 (1H, *s* (*br*), H-2'), 7.18 (1H, *dd*, J_1 = 8.5, J_2 = 2 Hz, H-6'); ^{13}C NMR: see Table 1; MS m/z (rel. int.): 280 (100) $[\text{M}]^+$, 145 (9), 132 (10), 131 (9), 129 (5), 128 (9), 117 (5), 115 (9), 107 (16), 91 (5), 77 (8), 43 (10).

Hydrogenation of 8. 16.5 mg compound 8 were hydrogenated at 14° on PtO_2 in MeOH soln. CC (2 g silica gel; S-2) yielded 16.1 mg 9 as a colourless oil. TLC (S-1): R_f 0.41, violet with anisaldehyde; IR $\nu_{\max}$ cm^{-1} : 3590 (OH), 2960, 2935, 2875, 2840, 1610, 1595; UV $\lambda_{\max}$ (log ϵ): 226 (4.23), 278 (3.65), 283 (3.60);

+ NaOH: 230 (4.19, sh), 244 (3.99, sh), 278 (3.65), 283 (3.67), 300 (3.35 s); ^1H NMR: δ 0.92 (3H, *t* (*br*), J = 7.5 Hz, CH_3 -9'), 1.15 (3H, *d*, J = 7.0 Hz, CH_3 -9), 1.40–1.80 (2H, *m*, CH_2 -8'), 2.52 (2H, *t* (*br*), J = 8 Hz, CH_2 -7'), 2.70–3.00 (2H, *m*, CH_2 -7), 3.19–3.58 (1H, *m*, H-8), 3.75 (3H, *s*, MeO), 4.71 (1H, *s* (*br*), OH), 6.62–6.77 (3H, *m*), 6.89–7.03 (4H, *m*); ^{13}C NMR: see Table 1; MS m/z (rel. int.): 284 (11) $[\text{M}]^+$, 178 (16), 177 (100), 133 (8), 107 (7).

Acetylation of 8. 16.4 mg compound 8 were acetylated using $\text{Ac}_2\text{O}-\text{C}_5\text{H}_5\text{N}$. CC (3 g silica gel, S-1) yielded 16 mg of 10 as a colourless oil. TLC (S-1): R_f 0.39, violet with anisaldehyde; IR $\nu_{\max}$ cm^{-1} : 2960, 2940, 2920, 2860, 2840, 1750, (C=O), 1630, 1600; UV $\lambda_{\max}$ (log ϵ): 220 (4.48), 262 (4.45), 295 (3.94 s); ^1H NMR: δ 1.82 (3H, *d* (*br*), J = 5 Hz, CH_3 -9'), 2.25 (3H, *s*), 3.75 (2H, *s* (*br*), CH_2 -7), 3.80 (3H, *s*, MeO), 5.04 (1H, *m*, H_A -9), 5.10 (1H, *m*, H_B -9), 6.05 (1H, *dq*, J_1 = 16, J_2 = 5 Hz, H-8'), 6.31 (1H, *d* (*br*), J = 16 Hz, H-7'), 6.71–7.23 (7H); ^{13}C NMR: see Table 1; MS m/z (rel. int.): 322 (91) $[\text{M}]^+$, 281 (27), 280 (100), 173 (11), 148 (7), 145 (14), 132 (6), 131 (11), 129 (11), 128 (18), 115 (16), 107 (23), 91 (10), 77, (9), 43 (51).

Table 1. ^{13}C NMR data for hermosillol (8), tetrahydro-hermosillol (9) and hermosillol acetate (10)

C	8	9	10
1	132.05 ^a	135.13 ^a	137.51
2	130.26	130.28	130.04
3	115.01	114.84	121.04
4	153.60	153.55	149.02 ^a
5	115.01	114.84	121.04
6	130.26	130.28	130.04
7	41.82	42.76	42.11
8	148.56	34.37	148.05 ^a
9	115.46	19.47	115.98
1'	130.32 ^b	133.80 ^a	130.36 ^b
2'	130.65 ^b	127.16 ^b	130.77 ^b
3'	125.70 ^c	134.67 ^a	125.89 ^c
4'	155.59	155.17	155.72
5'	111.04	110.73	111.02
6'	127.66 ^c	126.35 ^b	127.68 ^c
7'	131.98 ^a	37.48	131.85
8'	123.56	24.75	123.64
9'	18.21	13.75	18.22
CH ₃ O	55.61	55.66	55.60
C=O			169.23
CH ₃			21.01

a, b and c might be interchanged.

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REFERENCES

1. Achenbach, H., Groß, J., Dominguez, X. A., Cano, G., Star, J. V., Brussolo, L. d. C., Muñoz, G., Salgado, F. and López, L. (1987) *Phytochemistry* **26**, 1159.
2. *Urania Pflanzenreich, Höhere Pflanzen I* (1971) Urania Verlag Leipzig, Jena, Berlin.
3. Shreve, F. and Wiggins, I. (1964) *Vegetation and Flora of the Sonora Desert*, Vol. 1, 733, Stanford Univ. Press, Stanford.
4. Martinez, M. (1959) *Las Plantas Medicinales de Mexico*, 4th edn, p. 90. Ediciones Botas, Mexico.
5. Engler, A. (1964) *Syllabus der Pflanzenfamilien*, Vol. II. Gebr. Borntraeger, Berlin, Nikolasssee.
6. Hayashi, T. and Thomson, R. H. (1975) *Phytochemistry* **14**, 1085.
7. Bowden, B. F., Ritchie, E. and Taylor, W. C. (1972) *Aust. J. Chem.* **25**, 2659.
8. Read, R. W. and Taylor, W. C. (1979) *Aust. J. Chem.* **32**, 2317.
9. Stahl, E. and Ittel, I. (1981) *Planta Med.* **42**, 144.
10. Agrawal, P. K. and Thakur, R. S. (1985) *Magn. Reson. Chem.* **23**, 389.
11. Wenkert, E., Gottlieb, H. E., Gottlieb, O. R., Pereira, M. O. S. and Formiga, M. D. (1976) *Phytochemistry* **15**, 1547.
12. Diaz, P. P., Yoshida, M. and Gottlieb, O. R. (1980) *Phytochemistry* **19**, 285.
13. Takahashi, S. and Ogiso, A. (1970) *Chem. Pharm. Bull. (Japan)* **18**, 100.
14. Stahl, E. (1967) *Dünnschichtchromatographie*, p. 817. Springer Verlag, Berlin, Heidelberg, New York.